INCREASED 4-ENOYL-CoA REDUCTASE ACTIVITY IN LIVER MITOCHONDRIA OF RATS FED HIGH-FAT DIETS AND ITS EFFECT ON FATTY ACID OXIDATION AND THE INHIBITORY ACTION OF PENT-4-ENOATE

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1. Introduction

Administration of clofibrate to rats protects hepatic fatty acid oxidation against inhibition by hypoglycin [1] and pent-4-enoate [1,2]. The inhibitory action of pent-4-enoate is related with the accumulation of pent-2,4-dienoyl-CoA or a subsequent β -oxidation metabolite in the mitochondria [2,3]. Apparently, clofibrate prevents this accumulation by increasing the activity of 4-enoyl-CoA reductase, or rather 2,4-dienoyl-CoA reductase, since the enzyme is most active with 2,4-dienoyl-CoA esters [2,4].

Here we show that partially hydrogenated marine oils in the diet also lead to increased 4-enoyl-CoA reductase activity and concomitantly prevent inhibition of fatty acid oxidation by pent-4-enoate.

2. Methods

Male Wistar rats (150 g) were fed either a normal standard pellet diet or semisynthetic high fat diets for 15–18 days. These diets have been described in [5] and were modified to contain either 15% by wt. of partially hydrogenated marine oils or soya-bean oil.

Hepatocytes were prepared according to [6] except that Krebs-Henseleit bicarbonate buffer with 0.5 mM CaCl₂ was used as suspension medium and incubation medium which also contained 0.5 mM fatty acid free albumine and 1 mM [U-¹⁴C] palmitate.

NADPH-dependent 4-enoyl-CoA reductase was assayed as in [4] in the particle-free supernatant of isolated mitochondria treated with 0.2% deoxycholate.

Fatty acyl-CoA esters were synthesized and char-

acterized as in [7]. Pent-2,4-dienoate was prepared according to [8].

3. Results

Table 1 shows that the activity of 4-enoyl-CoA reductase with 2,4-dienoyl-CoA esters as substrates were about doubled in mitochondria from rats fed partially hydrogenated marine oils, as compared with those from normally fed rats. Also soya-bean oil diet led to an increase in the activity of this enzyme, but

Table 1
The activity of 4-enoyl-CoA reductase in liver mitochondria from rats fed either a normal pellet diet or diets containing partially hydrogenated marine oils or soya-bean oil

Diet	Substrate	No. obs.	Activity of 4-enoyl-CoA reductase
Normal	Sorboyl-CoA	8	27.4 ± 1.8
Marine oils	Sorboyl-CoA	7	60.5 ± 3.2
Soya-bean oil	Sorboyl-CoA	4	39.4 ± 1.6
Normal	Pent-2,4-dienoyl-CoA	3	46.1 ± 5.9
Marine oils	Pent-2,4-dienoyl-CoA	3	89.5 ± 8.5

The results are given as nmol NADPH oxidized . min⁻¹ . mg soluble mitochondrial protein⁻¹ (means ± SEM). Sorboyl-CoA and pent-2,4-dienoyl-CoA was 0.1 mM

Table 2
The oxidation of [U-14C] palmitate in hepatocytes from rats fed either a normal pellet diet or a diet containing partially hydrogenated marine oils in the presence and absence of pent-4-enoate, antimycin A or rotenone

Inhibitor	Normal rats		Rats fed marine oils	
None	20.1	23.3	55.7	64.9
Pent-4-enoate (2 mM)	6.0	9.1	38.2	45.4
	(30)	(39)	(69)	(70)
Antimycin A (2 μM)	6.3	6.8	12.0	12.5
	(31)	(29)	(22)	(19)
Rotenone (30 µM)	13.3	14.6	26.5	34.0
	(66)	(63)	(48)	(52)

The results are given as palmitate recovered as acid-soluble radioactive products (nmol. mg protein⁻¹. 30 min⁻¹), and (in parenthesis) as palmitate oxidized in % of oxidation in the absence of inhibitor. The parallels represent results obtained with hepatocytes from two different animals

less pronounced (table 1). We had used sorboyl-CoA as substrate since pent-2,4-dienoyl-CoA was not available. As is evident from table 1, the enzyme was even more active with pent-2,4-dienoyl-CoA. The apparent $K_{\rm m}$ -value with both this substrate and sorboyl-CoA was 22 μ M.

As shown in table 2, feeding partially hydrogenated marine oils stimulated palmitate oxidation and protected specifically against inhibition by pent-4-enoate.

4. Discussion

Increased peroxisomal β -oxidation in liver cells seems to be a common effect of clofibrate and high fat diets [9-11]. Feeding a clofibrate diet also leads to a 4-5-fold increase in 2,4-dienoyl-CoA reductase activity of the liver mitochondria and a complete abolition of the inhibitory action of pent-4-enoate [2]. Now we have found that high fat diets, especially with hydrogenated marine oils, also increase the activity of the reductase (table 1) and give a partial protection against pent-4-enoate (table 2). Compared with low-

fat diet or normal pellet diet, peroxisomal β -oxidation and 2,4-dienoyl-CoA reduction are increased 4-7-fold by clofibrate [2,12], 2-2.4-fold by the marine oil diet (table 1) [11] and 1.4-fold by the soya-bean oil diet (table 1) [11]. Thus, high fat diets and clofibrate have several common effects.

NADPH-dependent reduction of deca-2,4-dienoyl-CoA may be a step in the degradation of linoleic acid [4]. Our results support that 4-enoyl-CoA reductase (2,4-dienoyl-CoA reductase) participates in the degradation of unsaturated fatty acids and that changes in its activity is part of an adaptation to high fat diets.

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References

- Van Hoof, F., Hue, L. and Sherratt, H. S. A. (1979) Biochem, Soc. Trans. 7, 163-165.
- [2] Borrebaek, B., Osmundsen, H. and Bremer, J. (1980) Biochem. Biophys. Res. Commun. 93, 1173-1179.
- [3] Holland, P. C. and Sherratt, H. S. A. (1973) Biochem.J. 136, 157-171.
- [4] Kunau, W. H. and Dommes, P. (1978) Eur. J. Biochem. 91,533-544.
- [5] Thomassen, M. S., Strøm, E., Christiansen, E. N. and Norum, K. R. (1979) Lipids 14, 58-65.
- [6] Seglen, P. O. (1973) Exp. Cell Res. 82, 391-398.
- [7] Osmundsen, H., Neat, C. E. and Norum, K. R. (1979) FEBS Lett. 99, 292-296.
- [8] Muskat, I. E., Becker, B. C. and Lowenstein, J. S. (1930) J. Am. Chem. Soc. 52, 326-332.
- [9] Christiansen, R. Z. (1978) Biochim. Biophys. Acta 530, 314-324.
- [10] Christiansen, R. Z., Christiansen, E. N. and Bremer, J. (1979) Biochim. Biophys. Acta 573, 417-429.
- [11] Neat, C. E., Thomassen, M. S. and Osmundsen, H. (1980) Biochem. J. 186, 369-371.
- [12] Lazarow, P. B. and DeDuve, C. (1976) Proc. Natl. Acad. Sci. USA 73, 2043-2046.